

## Impacts of cooling and cryopreservation on human sperm capacitation, as measured by Cap-Score<sup>™</sup>

## Abstract

Introduction: Studies with fresh human sperm have shown that G<sub>M1</sub> localization patterns (Cap-Score<sup>™</sup>) quantify capacitation status. Using an outcome of clinical pregnancy, Cap-Score prospectively predicted a man's fertility and determined his probability of generating a pregnancy. Here, we evaluate the impacts of cooling and cryopreservation/thawing on capacitation using Cap-Score.

Methods: Semen was collected, liquefied and split into control and experimental treatments. Control samples were processed normally for Cap-Score. For the cooling experiments, samples were extended in TEST Yolk Buffer (TYB) and cooled overnight in a Styrofoam box with a cold pack (n=5). For the cryopreservation experiments, samples were frozen in TYB with glycerol (Cryo; n=10). After storage in LN<sub>2</sub>, the samples were thawed at 37°C for 3 min, mixed and then placed back into the water bath for another 3 min. Post-treatment, samples were washed, exposed to non-capacitating (NC) or capacitating (Cap) conditions, incubated for 3 hrs. and then Cap-Score was determined after an overnight fix.

Results: An increase was observed in the control Cap when compared to the control NC treatment in the cooling experiment (40±4 vs 24±4%; p<0.01). There was no difference between the control Cap and the experimental Cap with cooled sperm (40±4 vs 40±2; p=0.87). In the cryopreservation experiment, an increase was again seen in the control Cap over the control NC (33±3 vs 19±2; p<0.01). Cap-Score was unchanged for Cryo Cap when compared to control CAP (34±1% vs 33±3%; p=0.75). No difference was observed between the Cryo NC and Cryo Cap (33±3 vs 34±1; p=0.82). The Cryo NC was greater than the control NC (33±3 vs 19±2%; p<0.01).

Conclusion: Despite exposure to TYB or TYB with glycerol, the Cap-Score male fertility assay could still be performed. Semen extension in TYB and overnight maintenance at reduced temperature had no detectable impact on Cap-Score. In contrast, cryopreservation/thawing in TYB with glycerol induced capacitation-like membrane changes in sperm incubated under non-capacitating conditions, supporting reports in the literature of the "cryocapacitation" phenomenon. However, no differences were observed in Cap-Score between fresh sperm or sperm after freezing/thawing and then incubation with stimuli for capacitation. Identification of impacts on capacitation could optimize protocols intended to preserve male fertility as well as improve IUI and IVF outcomes.

## Introduction

Sperm must mature functionally in the process of capacitation to become able to fertilize. Capacitation depends on membrane lipid changes, and can be assessed by redistribution of the ganglioside G<sub>M1</sub>, the basis of the Cap-Score<sup>™</sup> male fertility assay. Cap-Score functionally assesses male fertility and was prospectively shown to predict pregnancy. Here, we determined the impact of cryopreservation/thawing and cooling using Cap-Score.



Figure 1. The effects of semen extension and cooling on Cap-Score<sup>™</sup>. Five samples were collected and split into control and chilled treatments (incubated with TYB and chilled overnight). Cap-Score was determined for control and chilled samples that were incubated with (Cap) or without capacitation stimuli (NC). The bar graph shows average Cap-Scores (yaxis) for each treatment (x-axis). An increase was observed in the Control-Cap when compared to the Control-NC treatment (40±4 vs 24±4%; p<0.01). There was no difference between the Control-Cap and the Chilled-Cap treatments ( $40\pm4$  vs  $40\pm2$ ; p=0.87).

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- Being able to identify impacts on capacitation could assist in the optimization of protocols intended to preserve male fertility and improve IUI and IVF outcomes.

